



Review

# Cannabidiol for Pain Treatment: Focus on Pharmacology and Mechanism of Action

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Abstract: Cannabis has a long history of medical use. Although there are many cannabinoids present in cannabis,  $\Delta 9$  tetrahydrocannabinol ( $\Delta 9$ -THC) and cannabidiol (CBD) are the two components found in the highest concentrations. CBD itself does not produce typical behavioral cannabimimetic effects and was thought not to be responsible for psychotropic effects of cannabis. Numerous anecdotal findings testify to the therapeutic effects of CBD, which in some cases were further supported by research findings. However, data regarding CBD's mechanism of action and therapeutic potential are abundant and omnifarious. Therefore, we review the basic research regarding molecular mechanism of CBD's action with particular focus on its analgesic potential. Moreover, this article describes the detailed analgesic and anti-inflammatory effects of CBD in various models, including neuropathic pain, inflammatory pain, osteoarthritis and others. The dose and route of the administration-dependent effect of CBD, on the reduction in pain, hyperalgesia or allodynia, as well as the production of pro and anti-inflammatory cytokines, were described depending on the disease model. The clinical applications of CBD-containing drugs are also mentioned. The data presented herein unravel what is known about CBD's pharmacodynamics and analgesic effects to provide the reader with current state-of-art knowledge regarding CBD's action and future perspectives for research.

**Keywords:** CBD; network pharmacology; pain; neuropathic; osteoarthritis

## 1. Introduction

Cannabis sativa L. has a long history of medical use with some of the earliest records found in Egyptian medical papyrus dating circa 1550 BC [1]. Although there are many cannabinoids present in Cannabis sp.,  $\Delta^9$  tetrahydrocannabinol ( $\Delta 9$ -THC) and cannabidiol (CBD) are the two components found at the highest concentrations (for review of complex interactions between phytocannabinoids [2]). Δ9-THC is the primary psychoactive ingredient of cannabis with an established partial agonist activity on both of the canonical cannabinoid receptors, CB1 and CB2. Therefore,  $\Delta 9$ -THC has been a natural subject of thousands of studies concerning pharmacology, toxicity and the therapeutic potential of cannabinoids in the intervening years. This research led to the discovery of the endocannabinoid system, which consists of G-protein coupled (GPCR) cannabinoid receptors and their endogenous ligands, endocannabinoids, anandamide (AEA) and 2-arachidonoylglycerol (2AG). Endocannabinoids are lipid compounds that are metabolized in multiple enzymatic pathways involving specific enzymes, such as fatty acid amide hydrolase (FAAH), monoacylglycerol lipase or alternative pathways involving non-specific enzymes, often interlinked with inflammatory mediators, such as cyclooxygenases or lipoxygenases (for review of endocannabinoid system metabolism pathways [3]). This interaction between metabolic pathways occurs due to the structural similarity between prostaglandins and endocannabinoids. However, the primary effects of endocannabinoids depend upon GPCR activation

and the type of  $\alpha$  subunit coupled with the receptor. CB1 and CB2 are primary couple with the  $G\alpha i/o$  subunit, which inhibits adenylate cyclase activity and therefore lowers intracellular  $Ca^{2+}$  concentration. Despite the specificity of the endocannabinoid mechanism of action, complex metabolic interactions between endocannabinoid system and inflammatory mediators need to be taken into consideration as well when studying the effects of various cannabinoid-based drugs.

Although isolated earlier [4] than  $\Delta 9$ -THC, CBD has remained a more elusive and poorly studied substance, because CBD itself does not produce typical behavioral cannabimimetic effects, and was thought not to be responsible for psychotropic effects of cannabis [5]. The complicated legal status of CBD, throughout the world, further restricted the research and professional knowledge about the therapeutic potential of this compound. In spite of the limitations, numerous anecdotal findings testify to the therapeutic effects of CBD, including anticonvulsant, antipsychotic, anxiolytic, neuroprotective and sleep-promoting effects, which are further supported by research [6]. Initial clinical evidence suggests that CBD possesses a desirable safety profile [7], while numerous preclinical findings present anti-inflammatory effects of CBD [8–12]. However, the pharmacodynamics of CBD have been difficult to elucidate. Initial reports demonstrated that CBD competes poorly with cannabinoid ligands at the orthosteric site of cannabinoid receptors [13], leading to the conclusion that any action of CBD occurs independent of cannabinoid receptors. Further studies revealed this conclusion as partially true. Indeed, CBD directly interacts with various receptors, enzymes and ion channels; however, it was also shown to directly and indirectly interact with the endocannabinoid system [14].

Taken together, these initial findings imply that CBD represents a promising phytocannabinoid-based treatment option. However, data regarding its mechanism of action and therapeutic potential are abundant and omnifarious. Therefore, we review the basic research regarding the molecular mechanisms of CBD's actions with particular focus on its analgesic potential. The data presented herein unravel what is known about CBD's pharmacodynamics and analgesic effects to provide readers with current state-of-art knowledge regarding CBD's action and future perspectives for research. A summary of the data discussed herein will be shared as Supplementary Materials.

## 2. Mechanism of Action

# 2.1. GPCRs

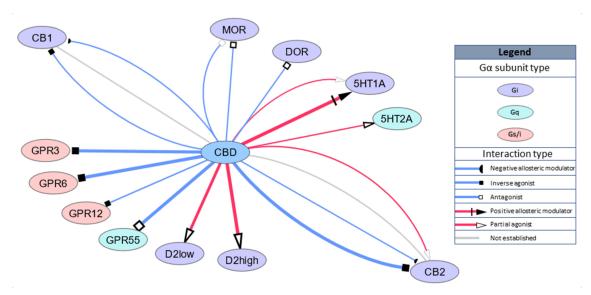
As mentioned above, CBD's binding to the orthosteric site of cannabinoid receptors is weak, with most studies reporting a Ki in the micromolar range [14–18] with one exception presenting Ki = 34 nM and partial agonism of CBD of the human CB2 receptors in HEK293A cells [14]. Surprisingly, much lower doses of CBD, in the nanomolar range, were able to antagonize the effects elicited by orthosteric agonists of CB1 and CB2, CP55940 and R-(+)-WIN55212 [17]. Recent developments in pharmacological studies explained this phenomenon through negative allosteric modulation of the cannabinoid receptors [14,18]. Moreover, it should be noted that CBD coadministration in vitro skews the signaling bias of various cannabinoid ligands [19,20].

The current state of knowledge has exposed CBD's mechanism action as distinct from the endocannabinoid system. Studies often present CBD's effects to be mediated by the serotoninergic 5HT1a receptor (5HT1a), which, similar to cannabinoid receptors, is coupled to the Gi protein. Although CBD's binding to the orthosteric site of 5HT1a is relatively weak (Ki = 16 mM [21]), 100 nM CBD produced an upward shift in the log concentration response curve of the 5HT1a agonist 8-OH-DPAT that resulted in a statistically significant increase in the Emax [22], suggesting positive allosteric modulation of 5HT1a.

CBD also interacts with various orphan GPCRs (GPR). CBD is antagonist for GPR55 (EC50 = 445 nM) [23] coupled with  $G_{13}$  alpha protein, regulating actin cytoskeletal remodeling in cells during movement and migration [24]. GPR55 has also been shown by other methods to use  $G_q$ ,  $G_{12}$ , or  $G_{13}$  for signal transduction, resulting in downstream activation of RhoA and PLC [24–26]. This signaling mode is associated with temporal changes in cytoplasmic calcium, membrane-bound diacylglycerol,

and plasma membrane topology. GPR55 is widely expressed in the brain, but its physiological function is still unclear. CBD is also an inverse agonist for GPR3, GPR6 and GPR12, with EC50 values of 1220 nM, 180 nM and 10,000 nM, respectively [27,28]. GPR3, GPR6 and GPR12 are primarily responsible for neurite outgrowth, cell survival and proliferation but have also been implicated in neuropathic pain development [29].

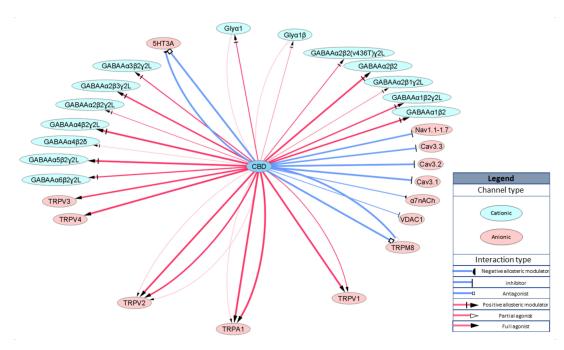
Studies have also revealed that CBD binds to other Gi-coupled receptors, namely, opioid receptors,  $\mu$ -opioid receptor (MOR) and  $\delta$ -opioid receptor (DOR) with Ki = 7000 nM and Ki = 10,000 nM, respectively, or Ki = 11 nM for D2 receptors with functional high-affinity for dopamine. A recent computational study revealed that the dopamine receptor D3 is a novel predicted target for CBD action [30]. GPCR targets are schematically summarized in Figure 1.



**Figure 1.** Schematic representation of cannabidiol (CBD) GPCR targets. Width of the edges (lines) represent relative affinity or EC/IC50 for the target (range 11–1000 nM).

# 2.2. Ionotropic Receptors

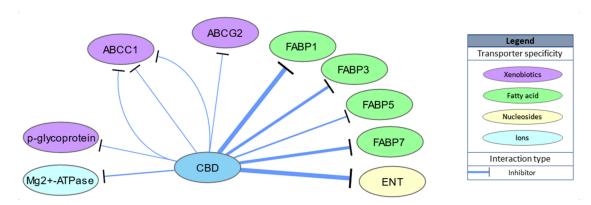
High activity for ionotropic receptors is a significant contributor to the physiological effects of CBD. Transient receptor potential (TRP) channels are a group of cationic ion channels primarily localized on the plasma membrane of numerous animal cell types. CBD activates TRPA1, TRPV1 and TRPV4 at low nanomolar concentrations [31–33] and TRPV2 at high micromolar concentrations [31,34,35]. Moreover, CBD is an antagonist for TRPM8 with an IC50 approximately 80 nM [31,32] and negative allosteric modulator for serotonin receptor 5HT3a (IC50 = 600 nM, [36]) and  $\alpha$ 7 nicotinic acetylcholine receptor (IC50 = 11,300 nM, [37]), which are also selective for positively charged ions. On the other hand, CBD is also a positive allosteric modulator in micromolar concentration ranges for anionic ligand-gated ion channels, such as GABA<sub>A</sub> [38] and glycine receptors (GlyRs) [39]. CBD also inhibits cationic currents through voltage gated calcium channels Cav3.1 and Cav3.2 with IC50 approximately 800 nM [40] and voltage-gated sodium channels and Cav3.3 with IC50 approximately 3000 nM [40,41]. CBD has also been shown to directly interact with voltage-dependent anion channel 1 (VDAC1), decreasing its conductance [42]. Ionotropic targets are schematically summarized in Figure 2.



**Figure 2.** Schematic representation of CBD ionotropic targets. Width of the edges (lines) represents relative EC/IC50 ranging between 60 and 20,000 nM.

#### 2.3. Transporters

CBD is also able to bind intracellular transporters of endocannabinoids, including fatty acid binding proteins 1, 3, 5 and 7 (Ki = 167, 1690, 1880 and 1520 nM, respectively), indirectly enhancing endocannabinoid actions through inhibition of anandamide uptake [43,44]. Indeed, CBD administration increases AEA levels in rat brains in an N-acyl phosphatidylethanolamine phospholipase D dependent manner [45] and in human serum [46]. Moreover, low nanomolar concentrations of CBD (IC50 = 124 nM) inhibit equilibrative nucleoside transporter (ENT) and adenosine uptake, which underlies both the anti-inflammatory and possibly sleep promoting effects of CBD [47,48]. CBD activity for other transporter proteins, such as multidrug resistance proteins (multidrug resistance-associated protein 1 (ABCC1), ATP-binding cassette super-family G member 2 (ABCG2) or P-glycoprotein) or Mg2+-ATPase, has also been studied; however, IC50 values above 3000 nM suggest these to be physiologically irrelevant. Transporter targets are schematically summarized in Figure 3.

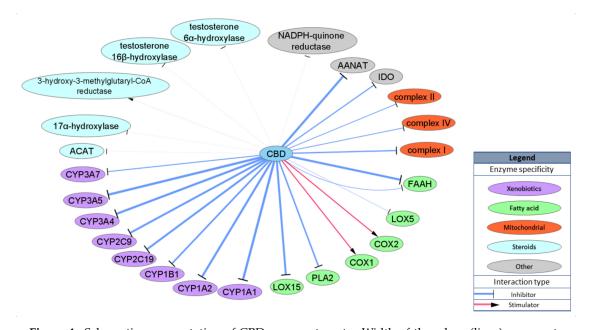


**Figure 3.** Schematic representation of CBD transporter targets. Width of the edges (lines) represents relative EC/IC50 ranging between 100 and 10,000 nM.

#### 2.4. Enzymes

Another important group of CBD targets is enzymes. The cytochrome P450 superfamily (CYPs) of enzymes is of particular interest as the interaction between CBD and CYPs may influence clearance

of various drugs, including very commonly used, non-steroidal anti-inflammatory drugs. It has been shown that CBD binds and inhibits activity of various CYPs, including CYP1B1, CYP2C19, CYP2C9, CYP3A4 and CYPC3A7, in physiologically relevant concentrations, with prominent IC50 values for CYP1A1 and CYP3A5 (IC50 = 77 and 195 nM, respectively). Due to the structural properties of CBD, it may also interact with various enzymes involved in lipid metabolism, most of which are involved in AEA metabolism. CBD inhibits rat FAAH activity with IC50 = 1520 nM [31], but the IC50 for the human isoform of FAAH is over 25,000 nM [33], which excludes FAAH as a possible target for CBD activity in humans. On the other hand, CBD inhibits alternative and nonspecific enzymes lipooxygenases (LOXs) involved in AEA degradation and production of inflammatory factors from arachidonic acid: LOX15 with IC50 = 2560 nM and LOX5 with IC50 = 73,730 nM [49]. Other studies have presented CBD as a stimulator of ovine COX1 and human recombinant COX2 with an IC50 approximately 10,000 nM [50] or as an inhibitor of phospholipase A2 (PLA2) from *Naja naja* venom with an IC50 = 6400 nM [51]. CBD has also been shown to inhibit mitochondrial complex I, II and IV but with very low potency (IC50 = 8200–19,100 nM) [52]. Moreover, CBD inhibits enzyme involved in the conversion of serotonin to melatonin, aralkylamine N-acetyltransferase (AANAT) with IC50 = 1000 nM [53], and an enzyme involved in tryptophan catabolism, indoleamine-pyrrole 2,3-dioxygenase (IDO) with IC50 = 8900 nM [54]. Few studies have examined CBD's interaction with enzymes involved in steroid metabolism such as acyl-CoA cholesterin acyltransferase (ACAT) or testosterone hydroxylase; however, either IC50 values were not specified or the concentration that was utilized extended beyond physiologically relevant values [55,56]. Enzymatic targets are schematically summarized in Figure 4.



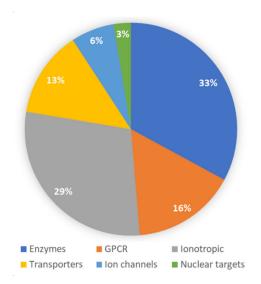
**Figure 4.** Schematic representation of CBD enzyme targets. Width of the edges (lines) represents relative EC/IC50 ranging between 77 and 30,000 nM.

#### 2.5. Nuclear Factors

One of the more important targets of CBD is the nuclear receptor peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ), especially in the context of inflammation. First, CBD has been shown to have low potency as a full agonist (EC50 = 20,100 nM) [57]; albeit a later study by Hedge et al., established an EC50 approximately 100 nM [58]. Moreover, studies by Juknat et al. revealed that nuclear factor erythroid-derived 2-like 2 (Nrf2) was responsible for CBD-mediated modulation of inflammatory gene expression patterns in activated microglia cells [59].

#### 2.6. Summary of Physiologically Relevant Pharmacological Targets

We identified a total of 76 different molecular targets of CBD. Among those, enzymes and ion channels/ionotropic receptors make up most of the targets. GPCRs and transporter proteins are less abundant targets of CBD, and there are only two different targets that are nuclear receptors. Among enzymes, CBD potently interacts with various CYP450 enzymes involved in drug metabolism, which raises concerns about possible drug interactions. Numerous targets among ion channels and ionotropic receptors might explain CBD's efficacy as a seizure controlling drug, but they may also contribute to the anti-nociceptive action of CBD. On the other hand, targets among GPCRs and transporters represent a link between CBD's actions and the endocannabinoid system because of CBD's high activity for the FABP proteins and allosteric modulation of cannabinoid receptors. The contribution of various protein types (enzyme, receptor, ion channel, etc.) is presented in Figure 5. We also analyzed the number of possible interactions with transmembrane receptors in relation to possible outcomes on cell membrane potential and cellular activity. Receptors coupled with Gi protein or anion-permeable channels were assumed to be inhibitory, whereas cation-permeable channels together with GPCRs coupled with Gq and possibly Gs were assumed to be stimulatory. This analysis revealed that almost 75% of direct CBD-receptor interactions should inhibit cell activity, whereas only 26% of interactions were able to stimulate cellular activity. More of the pharmacological effects of CBD, including the modulation of cytokine factors, cell function and reactive oxygen species may be found in the appendix table in the paper by McPartland et al., 2014 [60].



	Number of targets	% of all targets
total	76	
Enzymes	25	33
GPCR	12	16
Ionotropic	22	29
Transporters	10	13
Ion channels	5	7
Nuclear targets	2	3
Total GPCR types	12	
Gq	2	17
Gi	7	58
Gs/i	3	25
Ion permeability	27	
cationic	14	52
anionic	13	48
Total interactions	39	
stimulating	10	26
inhibitory	29	74

**Figure 5.** Pie chart showing the percentage of specific protein targets of CBD from a pool of all protein targets. Table presenting the number of identified CBD targets and percentage of specific  $G\alpha$  proteins coupled to GPCR receptors and ion permeability of both ion channels and ionotropic receptors. The total number of final effects on cell membrane potential was calculated based on the type of  $G\alpha$  proteins coupled to GPCR receptors targeted by CBD and ion permeability of those targets together with pharmacological action of CBD (for example, inhibition of anionic ion channel was counted as a stimulatory effect).

#### 3. The Analgesic Potential of CBD

Preclinical and clinical studies suggest a potential anti-nociceptive effect of CBD and CBD combined with other compounds in several pain-related diseases. Analgesic effects may vary depending on the dose and route of drug administration. The following section summarizes the newest findings in a various pain-related diseases and CBD effectiveness in pain management.

Several studies have reported that in healthy rodents submitted to a painful experience (e.g., tail-flick or paw pressure test) CBD administration may diminish the nociceptive experience. Moreover, depending on the experimental design, CBD may potentiate or antagonize  $\Delta 9$ -THC's

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effect. Intra-ventrolateral periaqueductal grey (PAG) microinjections of CBD or cannabichromene (CBC) resulted in a dose-dependent reduction in the ongoing activity of ON and OFF neurons in anaesthetized rats and caused analgesia, measured by the tail-flick test (effects were antagonized by selective antagonists of CB1, adenosine A1 and TRPA1, but not TRPV1 receptors) [61]. In turn, regarding the effects of CBD on  $\Delta 9$ -THC, Britch et al. reached inconsistent conclusions. In their study, CBD alone had no anti-nociceptive effect, however, it did increase animals' locomotor activity. When administered 15 min before  $\Delta 9$ -THC, CBD enhanced  $\Delta 9$ -THC-induced paw pressure, but not tail withdrawal, anti-nociception and enhanced rats' hypolocomotion (at low  $\Delta 9$ -THC dose). However, when administered 13 h or 15 min before  $\Delta 9$ -THC (in the lowest tested dose 1.8 mg/kg), CBD (in the highest tested dose of 30 mg/kg) had no effect on Δ9-THC's effect and inhibited Δ9-THC's metabolism (this effect was more prominent in females than in males) [62]. Greene et al. demonstrated that chronic daily CBD administration enhanced the development of tolerance to Δ9-THC-induced anti-nociception, likely due to CBD-induced inhibition of  $\Delta 9$ -THC metabolism or due to antagonism of  $\Delta 9$ -THC's effects after repeated treatment [63]. Cannabinoid-mediated neuromodulation may also be involved in transcutaneous electrical nerve stimulation (TENS)-mediated anti-nociception, a non-pharmacological therapy for the treatment of pain. In rats submitted to 10 or 150 Hz TENS, CBD and naloxone abolished the anti-nociceptive effects of both 10 and 150 Hz TENS [64].

## 3.1. Animal Models of Neuropathic Pain

Neuropathic pain arises from direct nerve damage or a disease and affects the somatosensory nervous system. Hyperalgesia, an excessive pain sensation, might be accompanied by allodynia, a painful sensation of neutral stimulus. Neuropathic pain is persistent and difficult to relieve; therefore, identification of novel, more effective therapeutic strategies is needed.

CBD exerts an analgesic effect in several neuropathic pain models. In a sciatic nerve injury mouse model, CBD-containing gelatine administered orally ad libitum significantly reduced allodynia up to 3 weeks post-surgery. Similar results were obtained in the Δ9-THC and morphine gelatine groups; however, tolerance to morphine developed after one week of treatment, which was not observed after CBD or  $\Delta 9$ -THC treatment. Moreover, the same study revealed no significant effect of CBD in hyperalgesia reduction, although an analgesic trend was observed [65]. In rats, after ligation of the L5 spinal nerve, CBD and its modified derivatives (dihydroxyl-CBD, and didesoxy-CBD) suppressed chronic neuropathic pain. The analgesic effect was correlated with cannabinoid potentiation of the  $\alpha$ 3 GlyRs but not with their binding affinity for CB1 and CB2 receptors. Furthermore, glycine currents in dorsal horn neurons in spinal cord slices from neuropathic rats were enhanced by cannabinoids [66]. In a mouse chronic neuropathic pain model involving foramen rotundum inflammatory constriction trigeminal infraorbital nerve injury, CBD administered orally in a peanut butter vehicle alleviated mechanical allodynia within 1 h and remained significant through 6 h [67]. CBD effectiveness was also demonstrated in a chronic constriction injury (CCI) animal model of neuropathic pain. Both oral and subcutaneous administration of CBD resulted in reduced surgery-induced mechanical and thermal allodynia [12,68]. Costa et al. also observed a reduction in the content of several pain mediators (prostaglandin E2 plasma concentration and paw tissue malondialdehyde, nitric oxide and glutathione-related enzymes levels) in response to 20 mg/kg oral administration of CBD. The anti-hyperalgesic effect was prevented by capsazepine (the vanilloid receptor antagonist) but not by rimonabant or SR144528 (CB1 and CB2 receptors antagonists, respectively) [12].  $\Delta 9$ -THC has stronger effects on pain reduction than CBD; nevertheless, side effects exclude its clinical use. However, Δ9-THC and CBD co-administration provided a 200-fold increase in low dose efficacy, with no side effects observed (only high co-administered doses caused side effects, similar to those seen with  $\Delta 9$ -THC alone) [68]. After spinal cord injury, CBD may act as an anti-inflammatory agent. Li et al. have shown that intraperitoneal (i.p.) CBD injections attenuated pro-inflammatory cytokine and chemokine invasion and prevented thermal sensitivity development [69]. In turn, in a peripheral sciatic nerve cuff neuropathic pain rat model, CBD, as well as  $\Delta 9$ -THC and CBD + Δ9-THC co-administration, reduced hypersensitivity and promoted beneficial changes in myelinated

Aβ mechanoreceptive fibers [70]. CBD also modulates serotoninergic transmission in dorsal raphe nucleus. Intravenous acute administration enhances the firing rate of 5-HT neurons in dorsal raphe nucleus of rats. This effect was blocked by 5HT1a and TRPV1 receptor antagonists (capsazepine and WAY100635, respectively). Repeated subcutaneous administration of CBD for 7 days enhances 5-HT firing through desensitization of 5HT1a receptors in healthy rats. In a spared nerve injury neuropathic pain rat model, repeated CBD injections diminished mechanical allodynia, anxiety-like behavior and normalized 5-HT activity. This effect was blocked by capsazepine and partially by WAY100635 [71].

In addition to surgically induced neuropathy, chronic pain can be triggered by chemical substance administration. For example, chemotherapeutic paclitaxel (PAC) might be responsible for peripheral neuropathy development in humans. In several studies of PAC-induced neuropathic pain, CBD exerted a positive effect against mechanical and thermal allodynia in mice and prevented mechanical sensitivity. The observed effect was reversed by the 5HT1a receptor antagonist WAY100635 but not by CB1 or CB2 receptor antagonists [72–74]. Furthermore, CBD and  $\Delta 9$ -THC co-administration enhanced the effect of low, ineffective doses of each drug alone [74]. In cisplatin-induced neuropathy, CBD reduced neuropathic pain in mice but did not prevent pain development, when administered 30 min prior to cisplatin [75]. Moreover, Pan et al. demonstrated that CBD prevents cisplatin-induced oxidative/nitrosative stress, inflammation and cell death in the kidney and improves renal function [11].

Type 1 diabetes is often associated with neuropathic pain development. In a rodent diabetes type 1 model, CBD delayed disease development and exerted a beneficial effect on pancreatic microcirculation in tested animals (reduction in leukocyte activation and increased functional capillary density) and attenuated myocardial dysfunction, cardiac fibrosis, oxidative/nitrosative stress, inflammation and cell death, while enhancing the ability of arteries to relax via enhanced production of vasodilator cyclooxygenase-1/2–derived products [50,76,77]. Intranasal and i.p. administration of CBD also attenuated neuropathic pain and inhibited elevation of microglial density and phosphorylated p38 mitogen-activated protein kinases [78]. Moreover, in streptozotocin-induced diabetic rats, acute i.p. CBD administration produced an anti-allodynic effect in the Von Frey test. This effect was mediated by serotoninergic system activation through 5HT1a receptors, since WAY100135, a selective 5HT1a receptor antagonist, but not CB1, CB2 or glycine antagonists, prevented the observed effect. When administered chronically, CBD reduced mechanical allodynia and prevented the decrease in serotonin levels in the spinal cord [79].

# 3.2. Inflammatory Pain

Inflammatory pain is caused by noxious stimuli that occur during the inflammatory or immune response. Under normal conditions, inflammation is a crucial protective mechanism, which plays an important role in the wound healing process. It is usually accompanied by redness, heat, swelling, pain/hypersensitivity, and loss of function. Nevertheless, in pathological conditions, inflammation may cause long-lasting pain through activation of sensory neurons [80].

In several inflammatory-induced chronic pain models, cannabinoids, including CBD, may exert an analgesic and anti-inflammatory effects. A study with complete Freund's adjuvant (CFA)-induced inflammatory pain in rodents revealed an important role for CBD and its modified derivatives in chronic pain attenuation [66]. In two additional murine models of induced inflammation, using 2.5% Croton oil in acetone topically applied to the ear or by i.p. injection of lipopolysaccharide (LPS), Verrico et al. demonstrated that CBD diminished serum levels of pro-inflammatory factors interleukin 6 (IL-6) and tumor necrosis factor  $\alpha$  (TNF $\alpha$ ) and increases levels of the anti-inflammatory cytokine interleukin 10 (IL-10). Similar results Verrico et al. were obtained in an in vitro model of inflammation in the same study [81]. On the other hand, Britch et al. demonstrated that twice daily i.p. administration of  $\Delta$ 9-THC for 3 days caused reduction in pain-related behavior in a CFA-induced inflammatory pain model but had little or no effect on changes in serum cytokines levels or paw edema. In turn, CBD i.p. administration had minimal effects on inflammatory pain but significantly reduced interleukin 1 $\beta$  (IL-1 $\beta$ ), IL-10, interferon  $\gamma$  (IFN- $\gamma$ ) levels and increased IL-6 levels [82]. This result suggests a more beneficial role for  $\Delta$ 9-THC compared to CBD in pain attenuation due

to the ambiguous results in inflammatory-related factor levels in the serum. Similar conclusions may be drawn from the work of Karmaus et al., where orally delivered CBD in corn oil enhanced LPS-induced pulmonary inflammation, augmented inflammatory cell infiltrate in bronchoalveolar lavage fluid and enhanced pro-inflammatory cytokine mRNA levels (TNFα, IL-6, interleukin-23 (IL-23), granulocyte colony stimulating factor (GCSF)) [83]. In another study, inflammatory pain was induced by carrageenan injection, and the effect of cannabidiolic acid (CBDA), CBD and Δ9-THC were investigated. I.p. administration of CBDA prior to carrageenan produced dose-dependent anti-hyperalgesia and anti-inflammatory effects. Similar effects were obtained with oral CBDA and  $\Delta$ 9-THC administration. A CB1 receptor antagonist, rimonabant, blocked the analgesic effect of  $\Delta$ 9-THC, while CBDA's effects were blocked by AMG9810 (a TRPV1 antagonist). Unfortunately, in this study, CBD did not reduce carrageenan-induced hyperalgesia. This result may suggest a more potent role for CBDA compared to CBD in inducing analgesic effects [84]. Moreno-Martet et al. investigated the role of Sativex-like (Sativex/Nabiximols is a cannabinoid drug containing 1:1 CBD and Δ9-THC) combined with a Δ9-tetrahydrocannabinol botanical drug substance (Δ9-THC-BDS) and cannabidiol-botanical drug substance (CBD-BDS) in an experimental autoimmune encephalitis (EAE) model of multiple sclerosis in mice. Results demonstrated that the Sativex-like combination and Δ9-THC-BDS alone improved neurological deficits in EAE animals and reduced the number and extent of cell aggregates in the spinal cord, derived from cell infiltration to the central nervous system. Rimonabant reversed the observed neurological benefits and reduced cell aggregates in the  $\Delta 9$ -THC-BDC-treated group. Notwithstanding, CBD-BDS alone did not provide these effects, merely delaying disease onset [85]. Moreover, CBD ameliorated the severity of experimental autoimmune encephalomyelitis in mice, diminishing axonal damage, inflammation and T-cell recruitment in the spinal cord of mice [10].

#### 3.3. Arthritis-Related Pain

Osteoarthritis (OA) is a degenerative joint disease primarily affecting people over 60 years old. Due to the increased ageing population and increased rates of obesity, OA has become a serious, global problem. There is no cure for this disease, and current therapy includes maintaining joint functionality through rehabilitation and losing weight, with pain alleviation using nonsteroidal anti-inflammatory drugs (NSAIDs) and opioids in more severe cases.

In preclinical models, CBD exerts an analgesic effect via different routes of administration. In a spontaneous canine OA model, CBD increased dogs' mobility and reduced pain, while no side effects were observed, Gamble et al. demonstrated increased alkaline phosphatase during CBD treatment [81,86]. In rodent OA models, disease is usually induced by intra-articular (i.a.) sodium monoiodoacetate (MIA) injection. In osteoarthritic rats, i.a. CBD administration dose-dependently decreased the joint afferent firing rate. Moreover, an increase in paw withdrawal threshold and weight bearing were observed. A TRPV1 antagonist, SB-366791, significantly inhibited the analgesic effect of CBD with respect to hind paw withdrawal threshold but did not have a significant effect on weight. CBD also reduced local inflammation by decreasing rolling and adherent leukocytes. The anti-rolling effect of CBD at 30 min was blocked by CB2 and TRPV1 antagonists but not a CB1 antagonist. The anti-adherence effect was blocked only by SB-366791. Prophylactic i.a. CBD administration prevented joint pain development and exerted a neuroprotective effect [87]. In a rat complete Freund's adjuvant-induced monoarthritic knee joint model, transdermal CBD gel applied for 4 days dose-dependently reduced joint swelling, limb posture scores, synovial membrane thickening and animals' pain. Furthermore, pro-inflammatory factors (Calcitonin gene-related peptide or OX42) were reduced in spinal cord and dorsal root ganglia (TNF $\alpha$ ) [9]. In a murine collagen type II-induced arthritis, daily i.p. or oral CBD administration reduced disease progression and blocked LPS-induced increases in serum  $TNF\alpha$ . In vitro culture of synovial cells collected from mice treated with CBD released decreased TNF $\alpha$  compared to cells collected from control animals. In vitro CBD also inhibited the release of reactive oxygen species by Zymosan-stimulated neutrophils [88]. The latest study by Lowin et al. showed that in human in vitro culture of rheumatoid arthritis synovial fibroblasts, CBD increases intracellular calcium levels, reduces cell viability, and IL-6/interleukin 8 (IL-8)/MMP-3

production. A TRPA1, but not TRPV1, antagonist reduced the effects of CBD. The effect was enhanced by TNF $\alpha$  pretreatment, which may suggest that CBD preferentially targets pro-inflammatory (activated) synovial fibroblasts, suggesting potential anti-arthritic activity [89]. Similar results from the work of Winklmayr et al. in an in vitro human chondrocytes culture showed that CBD concentrations greater than 4  $\mu$ M diminished cell viability and increased caspase 3/7 activity, elevated Ca<sup>2+</sup> and Extracellular signal-regulated kinases 1/2 phosphorylation and enhanced apoptotic cell population. This effect was mediated via the CB1 receptor, since AM251 inhibited Ca<sup>2+</sup> influx and reduced the toxic effects of CBD [90].

#### 3.4. Other Pain Models

Individual studies indicate a potential beneficial role for CBD in various disease animal models. Systemic (i.p.) injection of CBD reduced mechanical allodynia and reversed conditional place preference produced by peripheral nerve block in a rat model of incision pain. Moreover, direct rostral anterior cingulate cortex injection of CBD produced a similar, dose-dependent effect [91]. In a rat model of myofascial pain (induced by nerve growth factor intramuscular injection), intramuscular injection of CBD or cannabinol (CBN) decreased mechanical sensitization and increased the mechanical threshold of masseter muscle mechanoreceptors [92]. CBD may also enhance morphine-induced anti-nociception. In animal models in which N-methyl-D-aspartate (NMDA) receptor over activity plays a crucial role (opioid analgesia attenuation, NMDA-induced convulsive syndrome and ischemic stroke), intracerebroventricular (icv) injection of CBD enhanced morphine-evoked supraspinal anti-nociception, mitigated NMDA-induced convulsive syndrome and reduced the infarct size caused by permanent unilateral middle cerebral artery occlusion. This effect was absent in sigma-1 receptor ( $\sigma$ 1R) knock-out mice and was reduced by the σ1R agonists PRE084 and PPCC [93]. Neelakantan et al. demonstrated that distinct mechanisms of action may underlie the interactions between CBD and morphine in the different behavioral assays. Morphine alone diminished nociceptive responses in all three tested pain models (acetic acid-stimulated stretching, acetic acid-decreased operant responding for palatable food and hot plate thermal nociception), whereas CBD alone produced an anti-nociceptive effect only in the acetic acid-stimulated stretching model. Nevertheless, CBD and morphine combination produced synergistic effects in reversing acetic acid-stimulated stretching behavior, but subadditive effects in the hot plate thermal nociceptive assay and the acetic acid-decreased operant responding for palatable food assay [94]. In a 6-hydroxydopamine-induced Parkinson's disease mouse model, CBD exerted analgesic effects by increasing anandamide binding to the CB1 and TRPV1 receptors. URB597, a relatively selective FAAH inhibitor, potentiated the CBD effect, blockade of CB1 (by AM251) inhibited CBD effect, and TRPV1 receptor antagonism by capsazepine increased the anti-nociceptive effect of CBD [95]. In a corneal hyperalgesia mouse model, CBD produced anti-nociceptive effects and reduced neutrophil-infiltration in a wild-type and CB2 knock-out mice. The effect of CBD was blocked by a 5HT1a receptor antagonist, WAY100635, in wild-type mice, indicating the involvement of 5-HT1a and CB2 receptors in analgesic and anti-inflammatory actions [8]. However, Finn et al. proved that CBD alone did not reduce nociception in a rat model of persistent pain, in contrast to  $\Delta 9$ -THC, morphine and its combinations, and did not modulate  $\Delta 9$ -THC' anti-nociceptive effects [96].

## 3.5. Clinical Studies

The positive effects of CBD were confirmed many years ago [97–99], encouraging clinical trials. CBD was proven to be safe and cause only mild adverse effects in humans (e.g., ataxia, sedation, nausea, headache or decreased appetite) [100–103]. A transdermal CBD-containing gel in patients with peripheral neuropathic pain mitigated pain, as well as cold and itchy sensations [104]. CBD might be used clinically alone or in combination with other cannabinoids. Epidiolex, a pure CBD containing drug, is indicated for the treatment of seizures associated with Lennox Gastaut syndrome, Dravet syndrome, or tuberous sclerosis complex in patients 1 year of age and older. In treatment-resistant epilepsy, CBD reduced seizure frequency in both children and adults [105]. In Lennox-Gestaut syndrome, a rare and severe childhood-onset form of epilepsy, oral CBD reduced seizure frequency and improved

patients' overall conditions but triggered adverse effects (somnolence, decreased appetite, diarrhea; more severe in higher doses groups) [106,107]. CBD has shown similar results for the treatment of Dravet syndrome, a drug-resistant epilepsy that begins during the first year of life. CBD diminished major motor seizures and improved patients' overall condition [108,109].

In a study of chronic pain patients by Caparo et al., gels containing 15.7 mg CBD, 0.5 mg  $\Delta 9$ -THC, 0.3 mg cannabidivarin, 0.9 mg cannabidiolic acid, 0.8 mg cannabichrome and >1% botanical terpene blend caused reduced opioid use (in 53% of patients) and improved quality of life and sleep after 8 weeks of cannabinoid treatment [110]. However, in a study investigating three CBD- and  $\Delta 9$ -THC-containing drugs (Bedrocan 22.4 mg  $\Delta 9$ -THC + <1 mg CBD; Bediol 13.4 mg  $\Delta 9$ -THC + 17.8 mg CBD; Bedrolite 18.4 mg CBD + <1 mg  $\Delta 9$ -THC), none of the treatments had an effect greater than placebo on spontaneous or electrical pain responses. Drugs containing  $\Delta 9$ -THC increased the pressure pain threshold, while CBD increased  $\Delta 9$ -THC plasma concentrations [111].

The majority of clinical studies describe the efficacy of CBD and  $\Delta 9$ -THC co-administration, generally in doses of 2.5 mg CBD and 2.7 mg Δ9-THC in an oral mucosa spray, with treatment periods varying from one to several weeks. After treatment sessions, patients reported reduced pain, improved sleep quality, and reduced insomnia and fatigue [112-115]. One of the best studied CBD-containing registered products is Sativex<sup>®</sup> (Nabiximols), containing the abovementioned doses of CBD and  $\Delta 9$ -THC. Clinical studies have demonstrated its efficacy in pain attenuation in several diseases. It was proven to mitigate neuropathic pain and improve quality of life in patients suffering from several diseases, e.g., multiple sclerosis, cancer or rheumatoid arthritis [116–119]. Lichtman et al. proved that U.S. patients (but not patients from the rest of the world) experienced significant benefits from Nabiximols in cancer-related attenuation of pain. This effect might be due to lower doses of opioids taken at baseline or different distribution of cancer pain types [120]. In turn, Lynch et al. did not observe a significant reduction in pain intensity in chemotherapy-induced neuropathic pain compared to placebo. However, five of 16 patients reported a two-point or greater reduction in pain [121]. A study by Santoro et al. showed that in multiple sclerosis patients, treated or not with interferon-1β (IFN-1β), Sativex<sup>®</sup> decreased CNR2 expression levels in peripheral blood mononuclear cells (this effect was not observed in patients who were not treated with IFN-1β during the study) [122].

#### 4. Summary

CBD is a well-tolerated and safe natural compound exerting analgesic effects in various animal models of pain, as well as clinical studies. The referenced studies indicate a positive influence of CBD in treatment for various diseases in both pre-clinical and clinical trials. In the majority of animal studies, CBD has been demonstrated to exert analgesic effects, diminishing hyperalgesia and mechanical/thermal allodynia through various routes of administration. When co-administered with Δ9-THC, CBD may reduce the effective dose and diminish negative side effects of  $\Delta 9$ -THC [68,74]. However, some studies indicate no modulation of Δ9-THC's effects by CBD [96]. Moreover, CBD has the potential to act in an anti-inflammatory way; however; the effects are unclear. In the inflammatory-related pain modes, some research gives hope for an anti-inflammatory action of CBD, while others have reported opposing results. More detailed research is needed to clarify this effect; however, the inflammatory state is not always negative. In arthritic in vitro studies, CBD promoted chondrocyte and synoviocyte apoptosis but with stronger effects on the inflammatory-activated cells, which may be a positive result that indicates anti-arthritic activity [89,90]. The discussed studies indicate a positive influence of CBD on various diseases; nevertheless, animal studies cannot always be translated into human results. It is also important to remember that there are few to no studies on chronic CBD administration in healthy people. Despite the fact that there are trials on heathy volunteers that demonstrate CBD's safety and good tolerance, these studies do not examine prolonged use, which is common in chronically ill people. Patients suffering from chronic pain are often forced to take medications continuously for many years; it is not possible to test the effects of CBD on healthy people for such a long time [123,124]. Animal studies with chronic CBD treatment last several weeks, up to several months, which may

reflect the prolonged use in people, but the differences could not be excluded. In one animal study, Ignatowska-Jankowska et al. demonstrated that repeated CBD treatment may inhibit specific immunity by reducing T, B, T cytotoxic, and T helper cell numbers, while increasing the number of NK and NKT cells involved in nonspecific antiviral and antitumor immune response [125]. Chronic CBD treatment allows for long-term therapeutic effects to be achieved, without significant side effects or tolerance development. Especially important are prolonged anti-inflammatory effects, which may have an influence more directly on the cause of the pain development and therefore provide long-lasting analgesic effect. Acute CBD treatment may not be sufficient to combat the cause of the pain and provide an effect lasting up to several hours (depending on the disease model, dose and route of administration).

CBD is a substance of great therapeutic potential but is currently very understudied. Human CBD short-term studies illustrate low toxicity and mild adverse effects. However, it is unknown what effects will be caused by "CBD gummies" or "CBD water", which are increasingly offered by manufactures to a wide audience. CBD's pharmacology is not as clear as  $\Delta 9$ -THC, where the exact mechanism of action is known. Here, we still have much to learn.

Due to its complex pharmacological profile, effects observed from CBD administration vary and might be state-dependent. In vivo studies have revealed significant roles of various molecular targets in different models, namely, 5HT1a, CB1, CB2, TRPV1, α3 GlyRs, adenosine A1 and TRPA1. These targets were often not replicated throughout various models, suggesting the hypothesis of state-dependent effects of CBD. To further support this hypothesis, we emphasize CBD interactions with the endocannabinoid system. CBD may act as an indirect agonist of cannabinoid receptors through increased endocannabinoid tone [95], most likely through FABP inhibition [43,44], but on the other hand, it can directly antagonize the CB1 receptor [14]. Similarly, CBD activity for the CB2 receptor is very complex, including both partial agonism and negative allosteric modulation [14]. To complicate the issue even further, it is known that partial agonist action is dependent on receptor expression, density, and tonic activity of the system; therefore, it may vary in different tissues and under different conditions [126], further supporting our state-dependent hypothesis. Unsurprisingly, many of the abovementioned targets include ion channels, such as TRPV1 and TRPA1 receptors or  $\alpha$ 3 GlyRs. Furthermore, the role of adenosine receptors in the anti-inflammatory and anti-nociceptive effects of CBD is worth further study as CBD is a potent inhibitor of adenosine reuptake; therefore, adenosine receptors might be the node of CBD's action. Moreover, it would be interesting to see the role of other molecular targets of CBD involved in pain transmission, particularly dopamine D2 receptors or GPRs among GPCRs and, more importantly, various ionotropic targets, such as calcium or sodium channels, GABA receptors, 5HT3A or α7nACh receptors.

Compared to  $\Delta 9$ -THC, CBD has fewer unwanted side effects, and they are milder. These results provide hope for successful CBD use in the clinic in the future; however, more studies are required for precise elucidation of CBD's mechanisms of action. We hope that this review, together with data tables regarding CBD's pharmacology, may facilitate the understanding of CBD's potential in various pain conditions. Importantly, serious consideration should be applied to possible drug–drug interactions due to a significant proportion of CBD's interaction with cytochrome P450 isoforms.

**Supplementary Materials:** Supplementary materials can be found at <a href="http://www.mdpi.com/1422-0067/21/22/8870/s1">http://www.mdpi.com/1422-0067/21/22/8870/s1</a>. Materials include a table summarizing results regarding CBD's action in pain research and a CSV table containing CBD's pharmacological data. If you are willing to use the tables for further analysis, please cite this paper.

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#### **Abbreviations**

 $\Delta$ 9-THC  $\Delta$ 9 tetrahydrocannabinol

Δ9-THC-BDS Δ9-tetrahydrocannabinol botanical drug substance

o1R Sigma-1 receptor
 2-AG 2-arachidonoylglycerol
 5HT1a Serotonin 1A receptor

AANAT Aralkylamine N-acetyltransferase
ABCC1 Multidrug resistance-associated protein 1
ABCG2 ATP-binding cassette super-family G member 2

ACAT Acyl-CoA cholesterin acyltransferase

AEA Anandamide

CAMP Cyclic adenosine monophosphate

CB Cannabinoid receptor

CBD Cannabidiol
CBDA Cannabidiolic acid

CBD-BDS Cannabidiol-botanical drug substance

CBC Cannabichromene
CBN Cannabinol

CCI Chronic constriction injury
CFA Complete Freund's adjuvant
CHO Chinese hamster ovary cells
CYPs Cytochrome P450 superfamily

DOR  $\delta$ -opioid receptor

EAE Experimental autoimmune encephalitis
ENT Equilibrative nucleoside transporter
EROD Ethoxyresorufin-O-deethylase
FAAH Fatty acid amide hydrolase

GCSF Granulocyte colony stimulating factor

GlyRs Glycine receptors

GPCR G-protein coupled receptor

GPR Orphan G-protein coupled receptor

GTP Guanosine 5'-O-[gamma-thio]triphosphate

HEK293 Human embryonic kidney 293 cells

HPLC High-performance liquid chromatography
HTRF Homogeneous Time Resolved Fluorescence

i.a. Intra-articular

IDO Indoleamine-pyrrole 2,3-dioxygenase

 $\begin{array}{lll} \text{IFN-}\gamma & \text{Interferon-} \gamma \\ \text{IFN-}1\beta & \text{Interferon-}1\beta \\ \text{IL} & \text{Interleukin} \\ \text{i.p.} & \text{Intra-peritoneal} \\ \text{LOXs} & \text{Lipooxygenases} \\ \text{LPS} & \text{Lipopolysaccharide} \\ \text{MIA} & \text{Sodium monoiodoacetate} \end{array}$ 

MOR  $\mu$ -opioid receptor

NAM Negative allosteric modulator

NBD-stearate 12-N-methyl-(7-nitrobenz-2-oxa-1,3-diazo)aminostearic acid

NMDA *N*-methyl-D-aspartate receptor

NSAIDs Nonsteroidal anti-inflammatory drugs

OA Osteoarthritis PAC Paclitaxel PAG Periaqueductal grey

PAM Positive allosteric modulators
PBMC Peripheral blood mononuclear cell

PLA2 Phospholipase A2

 $PPR\gamma \hspace{1cm} Peroxisome \hspace{0.1cm} proliferator-activated \hspace{0.1cm} receptor \hspace{0.1cm} gamma$ 

TENS Transcutaneous electrical nerve stimulation

TNF $\alpha$  Tumor necrosis factor  $\alpha$ 

TRP Transient receptor potential channel VDAC1 Voltage-dependent anion channel 1

## References

- 1. Russo, E.B. History of Cannabis and Its Preparations in Saga, Science, and Sobriquet. *Chem. Biodivers.* **2007**, 4, 1614–1648. [CrossRef]
- 2. Ladha, K.S.; Ajrawat, P.; Yang, Y.; Clarke, H. Understanding the Medical Chemistry of the Cannabis Plant is Critical to Guiding Real World Clinical Evidence. *Molecules* **2020**, 25, 4042. [CrossRef] [PubMed]
- 3. Toczek, M.; Malinowska, B. Enhanced endocannabinoid tone as a potential target of pharmacotherapy. *Life Sci.* **2018**, *204*, 20–45. [CrossRef] [PubMed]
- 4. Adams, R.; Hunt, M. Structure of Cannabidiol, a Product Isolated from the Marihuana Extract of Minnesota Wild Hemp. I. *J. Am. Chem. Soc.* **1940**, *62*, 196–200. [CrossRef]
- 5. Mechoulam, R.; Shani, A.; Edery, H.; Grunfeld, Y. Chemical basis of hashish activity. *Science* **1970**, *169*, 611–612. [CrossRef]
- 6. Crippa, J.A.; Guimarães, F.S.; Campos, A.C.; Zuardi, A.W. Translational investigation of the therapeutic potential of cannabidiol (CBD): Toward a new age. *Front. Immunol.* **2018**, 9. [CrossRef]
- 7. Iffland, K.; Grotenhermen, F. An Update on Safety and Side Effects of Cannabidiol: A Review of Clinical Data and Relevant Animal Studies. *Cannabis Cannabinoid Res.* **2017**, *2*, 139–154. [CrossRef]
- 8. Thapa, D.; Cairns, E.A.; Szczesniak, A.M.; Toguri, J.T.; Caldwell, M.D.; Kelly, M.E.M. The Cannabinoids Δ8THC, CBD, and HU-308 Act via Distinct Receptors to Reduce Corneal Pain and Inflammation. *Cannabis Cannabinoid Res.* **2018**, *3*, 11–20. [CrossRef]
- 9. Hammell, D.C.; Zhang, L.P.; Ma, F.; Abshire, S.M.; McIlwrath, S.L.; Stinchcomb, A.L.; Westlund, K.N. Transdermal cannabidiol reduces inflammation and pain-related behaviours in a rat model of arthritis. *Eur. J. Pain* **2016**, *20*, 936–948. [CrossRef]
- 10. Kozela, E.; Lev, N.; Kaushansky, N.; Eilam, R.; Rimmerman, N.; Levy, R.; Ben-Nun, A.; Juknat, A.; Vogel, Z. Cannabidiol inhibits pathogenic T cells, decreases spinal microglial activation and ameliorates multiple sclerosis-like disease in C57BL/6 mice. *Br. J. Pharmacol.* **2011**, *163*, 1507–1519. [CrossRef]
- 11. Pan, H.; Mukhopadhyay, P.; Rajesh, M.; Patel, V.; Mukhopadhyay, B.; Gao, B.; Haskó, G.; Pacher, P. Cannabidiol attenuates cisplatin-Lnduced nephrotoxicity by decreasing oxidative/nitrosative stress, inflammation, and cell death. *J. Pharmacol. Exp. Ther.* **2009**, *328*, 708–714. [CrossRef] [PubMed]
- 12. Costa, B.; Trovato, A.E.; Comelli, F.; Giagnoni, G.; Colleoni, M. The non-psychoactive cannabis constituent cannabidiol is an orally effective therapeutic agent in rat chronic inflammatory and neuropathic pain. *Eur. J. Pharmacol.* **2007**, *556*, 75–83. [CrossRef] [PubMed]
- 13. Thomas, B.F.; Gilliam, A.F.; Burch, D.F.; Roche, M.J.; Seltzman, H.H. Comparative receptor binding analyses of cannabinoid agonists and antagonists. *J. Pharmacol. Exp. Ther.* **1998**, 285, 285–292. [PubMed]
- 14. Tham, M.; Yilmaz, O.; Alaverdashvili, M.; Kelly, M.E.M.; Denovan-Wright, E.M.; Laprairie, R.B. Allosteric and orthosteric pharmacology of cannabidiol and cannabidiol-dimethylheptyl at the type 1 and type 2 cannabinoid receptors. *Br. J. Pharmacol.* **2019**, *176*, 1455–1469. [CrossRef] [PubMed]
- 15. Showalter, V.M.; Compton, D.R.; Martin, B.R.; Abood, M.E. Evaluation of binding in a transfected cell line expressing a peripheral cannabinoid receptor (CB2): Identification of cannabinoid receptor subtype selective ligands. *J. Pharmacol. Exp. Ther.* **1996**, *278*, 989–999.
- 16. Thomas, A.; Ross, R.A.; Saha, B.; Mahadevan, A.; Razdan, R.K.; Pertwee, R.G. 6"-Azidohex-2"-yne-cannabidiol: A potential neutral, competitive cannabinoid CB1 receptor antagonist. *Eur. J. Pharmacol.* **2004**, *487*, 213–221. [CrossRef]

- 17. Thomas, A.; Baillie, G.L.; Phillips, A.M.; Razdan, R.K.; Ross, R.A.; Pertwee, R.G. Cannabidiol displays unexpectedly high potency as an antagonist of CB 1 and CB 2 receptor agonists in vitro. *Br. J. Pharmacol.* **2007**, *150*, 613–623. [CrossRef]
- 18. Martínez-Pinilla, E.; Varani, K.; Reyes-Resina, I.; Angelats, E.; Vincenzi, F.; Ferreiro-Vera, C.; Oyarzabal, J.; Canela, E.I.; Lanciego, J.L.; Nadal, X.; et al. Binding and signaling studies disclose a potential allosteric site for cannabidiol in cannabinoid CB2 receptors. *Front. Pharmacol.* **2017**, *8*, 744. [CrossRef]
- 19. Laprairie, R.B.; Bagher, A.M.; Kelly, M.E.M.; Denovan-Wright, E.M. Biased type 1 cannabinoid receptor signaling influences neuronal viability in a cell culture model of huntington diseases. *Mol. Pharmacol.* **2016**, 89, 364–375. [CrossRef]
- Navarro, G.; Reyes-Resina, I.; Rivas-Santisteban, R.; Sánchez de Medina, V.; Morales, P.; Casano, S.; Ferreiro-Vera, C.; Lillo, A.; Aguinaga, D.; Jagerovic, N.; et al. Cannabidiol skews biased agonism at cannabinoid CB1 and CB2 receptors with smaller effect in CB1-CB2 heteroreceptor complexes. *Biochem. Pharmacol.* 2018, 157, 148–158. [CrossRef]
- 21. Russo, E.B.; Burnett, A.; Hall, B.; Parker, K.K. Agonistic properties of cannabidiol at 5-HT1a receptors. *Neurochem. Res.* **2005**, *30*, 1037–1043. [CrossRef] [PubMed]
- 22. Rock, E.M.; Bolognini, D.; Limebeer, C.L.; Cascio, M.G.; Anavi-Goffer, S.; Fletcher, P.J.; Mechoulam, R.; Pertwee, R.G.; Parker, L.A. Cannabidiol, a nonpsychotropic component of cannabis, attenuates vomiting and nausea-like behaviour via indirect agonism of 5-HT 1A somatodendritic autoreceptors in the dorsal raphe nucleus. *Br. J. Pharmacol.* 2012, *165*, 2620–2634. [CrossRef] [PubMed]
- 23. Ryberg, E.; Larsson, N.; Sjögren, S.; Hjorth, S.; Hermansson, N.-O.; Leonova, J.; Elebring, T.; Nilsson, K.; Drmota, T.; Greasley, P.J. The orphan receptor GPR55 is a novel cannabinoid receptor. *Br. J. Pharmacol.* **2007**, 152, 1092–1101. [CrossRef] [PubMed]
- 24. Lauckner, J.E.; Jensen, J.B.; Chen, H.Y.; Lu, H.C.; Hille, B.; Mackie, K. GPR55 is a cannabinoid receptor that increases intracellular calcium and inhibits M current. *Proc. Natl. Acad. Sci. USA* **2008**, 105, 2699–2704. [CrossRef] [PubMed]
- 25. Henstidge, C.M.; Balenga, N.A.B.; Ford, L.A.; Ross, R.A.; Waldhoer, M.; Irving, A.J. The GPR55 ligand L-α-lysophosphatidylinositol promotes RhoA-dependent Ca 2+ signaling and NFAT activation. *FASEB J.* **2009**, 23, 183–193. [CrossRef] [PubMed]
- 26. Kapur, A.; Zhao, P.; Sharir, H.; Bai, Y.; Caron, M.G.; Barak, L.S.; Abood, M.E. Atypical responsiveness of the orphan receptor GPR55 to cannabinoid ligands. *J. Biol. Chem.* **2009**, *284*, 29817–29827. [CrossRef] [PubMed]
- 27. Laun, A.S.; Song, Z.H. GPR3 and GPR6, novel molecular targets for cannabidiol. *Biochem. Biophys. Res. Commun.* 2017, 490, 17–21. [CrossRef]
- 28. Brown, K.J.; Laun, A.S.; Song, Z.H. Cannabidiol, a novel inverse agonist for GPR12. *Biochem. Biophys. Res. Commun.* **2017**, 493, 451–454. [CrossRef]
- 29. Laun, A.S.; Shrader, S.H.; Brown, K.J.; Song, Z.H. GPR3, GPR6, and GPR12 as novel molecular targets: Their biological functions and interaction with cannabidiol. *Acta Pharmacol. Sin.* **2019**, *40*, 300–308. [CrossRef]
- 30. Bian, Y.M.; He, X.B.; Jing, Y.K.; Wang, L.R.; Wang, J.M.; Xie, X.Q. Computational systems pharmacology analysis of cannabidiol: A combination of chemogenomics-knowledgebase network analysis and integrated in silico modeling and simulation. *Acta Pharmacol. Sin.* **2019**, *40*, 374–386. [CrossRef]
- 31. De Petrocellis, L.; Ligresti, A.; Moriello, A.S.; Allarà, M.; Bisogno, T.; Petrosino, S.; Stott, C.G.; Di Marzo, V. Effects of cannabinoids and cannabinoid-enriched Cannabis extracts on TRP channels and endocannabinoid metabolic enzymes. *Br. J. Pharmacol.* **2011**, *163*, 1479–1494. [CrossRef] [PubMed]
- 32. De Petrocellis, L.; Vellani, V.; Schiano-Moriello, A.; Marini, P.; Magherini, P.C.; Orlando, P.; Di Marzo, V. Plant-derived cannabinoids modulate the activity of transient receptor potential channels of ankyrin type-1 and melastatin type-8. *J. Pharmacol. Exp. Ther.* **2008**, 325, 1007–1015. [CrossRef] [PubMed]
- 33. Bisogno, T.; Hanuš, L.; De Petrocellis, L.; Tchilibon, S.; Ponde, D.E.; Brandi, I.; Moriello, A.S.; Davis, J.B.; Mechoulam, R.; Di Marzo, V. Molecular targets for cannabidiol and its synthetic analogues: Effect on vanilloid VR1 receptors and on the cellular uptake and enzymatic hydrolysis of anandamide. *Br. J. Pharmacol.* 2001, 134, 845–852. [CrossRef] [PubMed]
- 34. Qin, N.; Neeper, M.P.; Liu, Y.; Hutchinson, T.L.; Lubin, M.L.; Flores, C.M. TRPV2 is activated by cannabidiol and mediates CGRP release in cultured rat dorsal root ganglion neurons. *J. Neurosci.* **2008**, *28*, 6231–6238. [CrossRef]

- 35. Nabissi, M.; Morelli, M.B.; Santoni, M.; Santoni, G. Triggering of the TRPV2 channel by cannabidiol sensitizes glioblastoma cells to cytotoxic chemotherapeutic agents. *Carcinogenesis* **2013**, *34*, 48–57. [CrossRef]
- 36. Yang, K.H.; Galadari, S.; Isaev, D.; Petroianu, G.; Shippenberg, T.S.; Oz, M. The nonpsychoactive cannabinoid cannabidiol inhibits 5- hydroxytryptamine3A receptor-mediated currents in xenopus laevis oocytes. *J. Pharmacol. Exp. Ther.* **2010**, 333, 547–554. [CrossRef]
- 37. Mahgoub, M.; Keun-Hang, S.Y.; Sydorenko, V.; Ashoor, A.; Kabbani, N.; Al Kury, L.; Sadek, B.; Howarth, C.F.; Isaev, D.; Galadari, S.; et al. Effects of cannabidiol on the function of α7-nicotinic acetylcholine receptors. *Eur. J. Pharmacol.* **2013**, 720, 310–319. [CrossRef]
- 38. Bakas, T.; van Nieuwenhuijzen, P.S.; Devenish, S.O.; McGregor, I.S.; Arnold, J.C.; Chebib, M. The direct actions of cannabidiol and 2-arachidonoyl glycerol at GABAA receptors. *Pharmacol. Res.* **2017**, *119*, 358–370. [CrossRef]
- 39. Ahrens, J.; Demir, R.; Leuwer, M.; De La Roche, J.; Krampfl, K.; Foadi, N.; Karst, M.; Haeseler, G. The nonpsychotropic cannabinoid cannabidiol modulates and directly activates alpha-1 and alpha-1-beta glycine receptor function. *Pharmacology* **2009**, *83*, 217–222. [CrossRef]
- 40. Ross, H.R.; Napier, I.; Connor, M. Inhibition of recombinant human T-type calcium channels by Δ9-tetrahydrocannabinol and cannabidiol. *J. Biol. Chem.* **2008**, *283*, 16124–16134. [CrossRef]
- 41. Ghovanloo, M.R.; Shuart, N.G.; Mezeyova, J.; Dean, R.A.; Ruben, P.C.; Goodchild, S.J. Inhibitory effects of cannabidiol on voltage-dependent sodium currents. *J. Biol. Chem.* **2019**, 293, 16546–16558. [CrossRef] [PubMed]
- 42. Rimmerman, N.; Ben-Hail, D.; Porat, Z.; Juknat, A.; Kozela, E.; Daniels, M.P.; Connelly, P.S.; Leishman, E.; Bradshaw, H.B.; Shoshan-Barmatz, V.; et al. Direct modulation of the outer mitochondrial membrane channel, voltage-dependent anion channel 1 (VDAC1) by cannabidiol: A novel mechanism for cannabinoid-induced cell death. *Cell Death Dis.* **2013**, *4*, e949. [CrossRef] [PubMed]
- 43. Elmes, M.W.; Kaczocha, M.; Berger, W.T.; Leung, K.; Ralph, B.P.; Wang, L.; Sweeney, J.M.; Miyauchi, J.T.; Tsirka, S.E.; Ojima, I.; et al. Fatty Acid-binding Proteins (FABPs) Are Intracellular Carriers for Δ<sup>9</sup>-Tetrahydrocannabinol (THC) and Cannabidiol (CBD). *J. Biol. Chem.* 2015, 290, 8711–8721. [CrossRef] [PubMed]
- 44. Huang, H.; McIntosh, A.L.; Martin, G.G.; Landrock, D.; Chung, S.; Landrock, K.K.; Dangott, L.J.; Li, S.; Kier, A.B.; Schroeder, F. FABP1: A Novel Hepatic Endocannabinoid and Cannabinoid Binding Protein. *Biochemistry* **2016**, *55*, 5243–5255. [CrossRef] [PubMed]
- 45. Leishman, E.; Manchanda, M.; Thelen, R.; Miller, S.; Mackie, K.; Bradshaw, H.B. Cannabidiol's Upregulation of N-acyl Ethanolamines in the Central Nervous System Requires N-acyl Phosphatidyl Ethanolamine-Specific Phospholipase D. *Cannabis Cannabinoid Res.* 2018, 3, 228–241. [CrossRef] [PubMed]
- 46. Leweke, F.M.; Piomelli, D.; Pahlisch, F.; Muhl, D.; Gerth, C.W.; Hoyer, C.; Klosterkötter, J.; Hellmich, M.; Koethe, D. Cannabidiol enhances anandamide signaling and alleviates psychotic symptoms of schizophrenia. *Transl. Psychiatry* **2012**, *2*, e94. [CrossRef] [PubMed]
- 47. Carrier, E.J.; Auchampach, J.A.; Hillard, C.J. Inhibition of an equilibrative nucleoside transporter by cannabidiol: A mechanism of cannabinoid immunosuppression. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 7895–7900. [CrossRef] [PubMed]
- 48. Mijangos-Moreno, S.; Poot-Aké, A.; Arankowsky-Sandoval, G.; Murillo-Rodríguez, E. Intrahypothalamic injection of cannabidiol increases the extracellular levels of adenosine in nucleus accumbens in rats. *Neurosci. Res.* **2014**, *84*, 60–63. [CrossRef]
- 49. Takeda, S.; Usami, N.; Yamamoto, I.; Watanabe, K. Cannabidiol-2',6'-dimethyl ether, a cannabidiol derivative, is a highly potent and selective 15-lipoxygenase inhibitor. *Drug Metab. Dispos.* **2009**, 37, 1733–1737. [CrossRef]
- 50. Wheal, A.J.; Cipriano, M.; Fowler, C.J.; Randall, M.D.; O'Sullivan, S.E. Cannabidiol improves vasorelaxation in Zucker diabetic fatty rats through cyclooxygenase activation. *J. Pharmacol. Exp. Ther.* **2014**, 351, 457–466. [CrossRef]
- 51. Evans, A.T.; Formukong, E.; Evans, F.J. Activation of phospholipase A2 by cannabinoids. Lack of correlation with CNS effects. *FEBS Lett.* **1987**, *211*, 119–122. [CrossRef]
- 52. Fišar, Z.; Singh, N.; Hroudová, J. Cannabinoid-induced changes in respiration of brain mitochondria. *Toxicol. Lett.* **2014**, 231, 62–71. [CrossRef] [PubMed]

- 53. Koch, M.; Dehghani, F.; Habazettl, I.; Schomerus, C.; Korf, H.W. Cannabinoids attenuate norepinephrine-induced melatonin biosynthesis in the rat pineal gland by reducing arylalkylamine N-acetyltransferase activity without involvement of cannabinoid receptors. *J. Neurochem.* 2006, 98, 267–278. [CrossRef] [PubMed]
- 54. Jenny, M.; Santer, E.; Pirich, E.; Schennach, H.; Fuchs, D. Δ9-Tetrahydrocannabinol and cannabidiol modulate mitogen-induced tryptophan degradation and neopterin formation in peripheral blood mononuclear cells in vitro. *J. Neuroimmunol.* 2009, 207, 75–82. [CrossRef]
- 55. Cornicelli, J.A.; Gilman, S.R.; Krom, B.A.; Kottke, B.A. Cannabinoids impair the formation of cholesteryl ester in cultured human cells. *Arteriosclerosis* **1981**, *1*, 449–454. [CrossRef]
- Watanabe, K.; Motoya, E.; Matsuzawa, N.; Funahashi, T.; Kimura, T.; Matsunaga, T.; Arizono, K.; Yamamoto, I. Marijuana extracts possess the effects like the endocrine disrupting chemicals. *Toxicology* 2005, 206, 471–478. [CrossRef]
- 57. Granja, A.G.; Carrillo-Salinas, F.; Pagani, A.; Gómez-Cañas, M.; Negri, R.; Navarrete, C.; Mecha, M.; Mestre, L.; Fiebich, B.L.; Cantarero, I.; et al. A cannabigerol quinone alleviates neuroinflammation in a chronic model of multiple sclerosis. *J. Neuroimmune Pharmacol.* **2012**, *7*, 1002–1016. [CrossRef]
- 58. Hegde, V.L.; Singh, U.P.; Nagarkatti, P.S.; Nagarkatti, M. Critical Role of Mast Cells and Peroxisome Proliferator–Activated Receptor  $\gamma$  in the Induction of Myeloid-Derived Suppressor Cells by Marijuana Cannabidiol In Vivo. *J. Immunol.* **2015**, *194*, 5211–5222. [CrossRef]
- 59. Juknat, A.; Pietr, M.; Kozela, E.; Rimmerman, N.; Levy, R.; Gao, F.; Coppola, G.; Geschwind, D.; Vogel, Z. Microarray and Pathway Analysis Reveal Distinct Mechanisms Underlying Cannabinoid-Mediated Modulation of LPS-Induced Activation of BV-2 Microglial Cells. *PLoS ONE* **2013**, *8*, e61462. [CrossRef]
- 60. McPartland, J.M.; Duncan, M.; Di Marzo, V.; Pertwee, R.G. Are cannabidiol and Δ9-tetrahydrocannabivarin negative modulators of the endocannabinoid system? A systematic review. *Br. J. Pharmacol.* **2015**, 172, 737–753. [CrossRef]
- 61. Maione, S.; Piscitelli, F.; Gatta, L.; Vita, D.; De Petrocellis, L.; Palazzo, E.; De Novellis, V.; Di Marzo, V. Non-psychoactive cannabinoids modulate the descending pathway of antinociception in anaesthetized rats through several mechanisms of action. *Br. J. Pharmacol.* **2011**, *162*, 584–596. [CrossRef] [PubMed]
- 62. Britch, S.C.; Wiley, J.L.; Yu, Z.; Clowers, B.H.; Craft, R.M. Cannabidiol-Δ9-tetrahydrocannabinol interactions on acute pain and locomotor activity. *Drug Alcohol Depend.* **2017**, *175*, 187–197. [CrossRef] [PubMed]
- 63. Greene, N.Z.; Wiley, J.L.; Yu, Z.; Clowers, B.H.; Craft, R.M. Cannabidiol modulation of antinociceptive tolerance to Δ9-tetrahydrocannabinol. *Psychopharmacology* **2018**, 235, 3289–3302. [CrossRef] [PubMed]
- 64. Gonçalves, T.C.T.; Londe, A.K.; Albano, R.I.P.; De Araújo Júnior, A.T.; De Aguiar Azeredo, M.; Biagioni, A.F.; Vasconcellos, T.H.F.; Dos Reis Ferreira, C.M.; Teixeira, D.G.; De Souza Crippa, J.A.; et al. Cannabidiol and endogenous opioid peptide-mediated mechanisms modulate antinociception induced by transcutaneous electrostimulation of the peripheral nervous system. *J. Neurol. Sci.* 2014, 347, 82–89. [CrossRef] [PubMed]
- 65. Abraham, A.D.; Leung, E.J.Y.; Wong, B.A.; Rivera, Z.M.G.; Kruse, L.C.; Clark, J.J.; Land, B.B. Orally consumed cannabinoids provide long-lasting relief of allodynia in a mouse model of chronic neuropathic pain. *Neuropsychopharmacology* **2020**, *45*, 1105–1114. [CrossRef] [PubMed]
- 66. Xiong, W.; Cui, T.; Cheng, K.; Yang, F.; Chen, S.R.; Willenbring, D.; Guan, Y.; Pan, H.L.; Ren, K.; Xu, Y.; et al. Cannabinoids suppress inflammatory and neuropathic pain by targeting α3 glycine receptors. *J. Exp. Med.* **2012**, 209, 1121–1134. [CrossRef] [PubMed]
- 67. Vigil, J.M.; Montera, M.A.; Pentkowski, N.S.; Diviant, J.P.; Orozco, J.; Ortiz, A.L.; Rael, L.J.; Westlund, K.N. The therapeutic effectiveness of full spectrum hemp oil using a chronic neuropathic pain model. *Life* **2020**, 10, 69. [CrossRef]
- 68. Casey, S.L.; Atwal, N.; Vaughan, C.W. Cannabis constituent synergy in a mouse neuropathic pain model. *Pain* **2017**, *158*, 2452–2460. [CrossRef]
- 69. Li, H.; Kong, W.; Chambers, C.R.; Yu, D.; Ganea, D.; Tuma, R.F.; Ward, S.J. The non-psychoactive phytocannabinoid cannabidiol (CBD) attenuates pro-inflammatory mediators, T cell infiltration, and thermal sensitivity following spinal cord injury in mice. *Cell. Immunol.* **2018**, 329, 1–9. [CrossRef]
- 70. Linher-Melville, K.; Zhu, Y.F.; Sidhu, J.; Parzei, N.; Shahid, A.; Seesankar, G.; Ma, D.; Wang, Z.; Zacal, N.; Sharma, M.; et al. Evaluation of the preclinical analgesic efficacy of naturally derived, orally administered oil forms of Δ9-tetrahydrocannabinol (THC), cannabidiol (CBD), and their 1:1 combination. *PLoS ONE* **2020**, 15, e0234176. [CrossRef]

- 71. De Gregorio, D.; McLaughlin, R.J.; Posa, L.; Ochoa-Sanchez, R.; Enns, J.; Lopez-Canul, M.; Aboud, M.; Maione, S.; Comai, S.; Gobbi, G. Cannabidiol modulates serotonergic transmission and reverses both allodynia and anxiety-like behavior in a model of neuropathic pain. *Pain* **2019**, *160*, 136–150. [CrossRef] [PubMed]
- 72. Ward, S.J.; Ramirez, M.D.; Neelakantan, H.; Walker, E.A. Cannabidiol prevents the development of cold and mechanical allodynia in paclitaxel-treated female c57bl6 mice. *Anesth. Analg.* **2011**, *113*, 947–950. [CrossRef] [PubMed]
- 73. Ward, S.J.; McAllister, S.D.; Kawamura, R.; Murase, R.; Neelakantan, H.; Walker, E.A. Cannabidiol inhibits paclitaxel-induced neuropathic pain through 5-HT 1A receptors without diminishing nervous system function or chemotherapy efficacy. *Br. J. Pharmacol.* **2014**, *171*, 636–645. [CrossRef] [PubMed]
- 74. King, K.M.; Myers, A.M.; Soroka-Monzo, A.J.; Tuma, R.F.; Tallarida, R.J.; Walker, E.A.; Ward, S.J. Single and combined effects of Δ9-tetrahydrocannabinol and cannabidiol in a mouse model of chemotherapy-induced neuropathic pain. *Br. J. Pharmacol.* **2017**, *174*, 2832–2841. [CrossRef]
- 75. Harris, H.M.; Sufka, K.J.; Gul, W.; Elsohly, M.A. Effects of Delta-9-Tetrahydrocannabinol and Cannabidiol on Cisplatin-Induced Neuropathy in Mice. *Planta Med.* **2016**, *82*, 1169–1172. [CrossRef]
- 76. Lehmann, C.; Fisher, N.B.; Tugwell, B.; Szczesniak, A.; Kelly, M.; Zhou, J. Experimental cannabidiol treatment reduces early pancreatic inflammation in type 1 diabetes. *Clin. Hemorheol. Microcirc.* **2016**, *64*, 655–662. [CrossRef]
- 77. Rajesh, M.; Mukhopadhyay, P.; Btkai, S.; Patel, V.; Saito, K.; Matsumoto, S.; Kashiwaya, Y.; Horvth, B.; Mukhopadhyay, B.; Becker, L.; et al. Cannabidiol attenuates cardiac dysfunction, oxidative stress, fibrosis, and inflammatory and cell death signaling pathways in diabetic cardiomyopathy. *J. Am. Coll. Cardiol.* **2010**, 56, 2115–2125. [CrossRef]
- 78. Toth, C.C.; Jedrzejewski, N.M.; Ellis, C.L.; Frey, W.H. Cannabinoid-Mediated Modulation of Neuropathic Pain and Microglial Accumulation in a Model of Murine Type I Diabetic Peripheral Neuropathic Pain. *Mol. Pain* 2010, 6. [CrossRef]
- 79. Jesus, C.H.A.; Redivo, D.D.B.; Gasparin, A.T.; Sotomaior, B.B.; de Carvalho, M.C.; Genaro, K.; Zuardi, A.W.; Hallak, J.E.C.; Crippa, J.A.; Zanoveli, J.M.; et al. Cannabidiol attenuates mechanical allodynia in streptozotocin-induced diabetic rats via serotonergic system activation through 5-HT1A receptors. *Brain Res.* 2019, 1715, 156–164. [CrossRef]
- 80. Vasko, M.R. I Ia Inhibitory Interneuron Definition. In *Inflammatory Pain*; Binder, M.D., Hirokawa, N., Windhorst, U., Eds.; Encyclopedia of Neuroscience; Springer: Berlin/Heidelberg, Germany, 2009.
- 81. Verrico, C.D.; Wesson, S.; Konduri, V.; Hofferek, C.J.; Vazquez-Perez, J.; Blair, E.; Dunner, K.; Salimpour, P.; Decker, W.K.; Halpert, M.M. A randomized, double-blind, placebo-controlled study of daily cannabidiol for the treatment of canine osteoarthritis pain. *Pain* **2020**, *161*, 2191–2202. [CrossRef]
- 82. Britch, S.C.; Goodman, A.G.; Wiley, J.L.; Pondelick, A.M.; Craft, R.M. Antinociceptive and Immune Effects of Delta-9-Tetrahydrocannabinol or Cannabidiol in Male Versus Female Rats with Persistent Inflammatory Pain. *J. Pharmacol. Exp. Ther.* **2020**, *373*, 416–428. [CrossRef] [PubMed]
- 83. Karmaus, P.W.F.; Wagner, J.G.; Harkema, J.R.; Kaminski, N.E.; Kaplan, B.L.F. Cannabidiol (CBD) enhances lipopolysaccharide (LPS)-induced pulmonary inflammation in C57BL/6 mice. *J. Immunotoxicol.* **2013**, 10, 321–328. [CrossRef] [PubMed]
- 84. Rock, E.M.; Limebeer, C.L.; Parker, L.A. Effect of cannabidiolic acid and Δ9-tetrahydrocannabinol on carrageenan-induced hyperalgesia and edema in a rodent model of inflammatory pain. *Psychopharmacology* **2018**, 235, 3259–3271. [CrossRef] [PubMed]
- 85. Moreno-Martet, M.; Feliú, A.; Espejo-Porras, F.; Mecha, M.; Carrillo-Salinas, F.J.; Fernández-Ruiz, J.; Guaza, C.; De Lago, E. The disease-modifying effects of a Sativex-like combination of phytocannabinoids in mice with experimental autoimmune encephalomyelitis are preferentially due to Δ9-tetrahydrocannabinol acting through CB1 receptors. *Mult. Scler. Relat. Disord.* **2015**, *4*, 505–511. [CrossRef]
- 86. Gamble, L.J.; Boesch, J.M.; Frye, C.W.; Schwark, W.S.; Mann, S.; Wolfe, L.; Brown, H.; Berthelsen, E.S.; Wakshlag, J.J. Pharmacokinetics, Safety, and Clinical Efficacy of Cannabidiol Treatment in Osteoarthritic Dogs. *Front. Vet. Sci.* **2018**, *5*, 165. [CrossRef]
- 87. Philpott, H.T.; O'Brien, M.; McDougall, J.J. Attenuation of early phase inflammation by cannabidiol prevents pain and nerve damage in rat osteoarthritis. *Pain* **2017**, *158*, 2442–2451. [CrossRef]

- 88. Malfait, A.M.; Gallily, R.; Sumariwalla, P.F.; Malik, A.S.; Andreakos, E.; Mechoulam, R.; Feldmann, M. The nonpsychoactive cannabis constituent cannabidiol is an oral anti-arthritic therapeutic in murine collagen-induced arthritis. *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 9561–9566. [CrossRef]
- 89. Lowin, T.; Tingting, R.; Zurmahr, J.; Classen, T.; Schneider, M.; Pongratz, G. Cannabidiol (CBD): A killer for inflammatory rheumatoid arthritis synovial fibroblasts. *Cell Death Dis.* **2020**, *11*, 1–11. [CrossRef]
- 90. Winklmayr, M.; Gaisberger, M.; Kittl, M.; Fuchs, J.; Ritter, M.; Jakab, M. Dose-Dependent Cannabidiol-Induced Elevation of Intracellular Calcium and Apoptosis in Human Articular Chondrocytes. *J. Orthop. Res.* **2019**, 37, 2540–2549. [CrossRef]
- 91. Genaro, K.; Fabris, D.; Arantes, A.L.F.; Zuardi, A.W.; Crippa, J.A.S.; Prado, W.A. Cannabidiol is a potential therapeutic for the affective-motivational dimension of incision pain in rats. *Front. Pharmacol.* **2017**, *8*, 391. [CrossRef]
- 92. Wong, H.; Cairns, B.E. Cannabidiol, cannabinol and their combinations act as peripheral analysis in a rat model of myofascial pain. *Arch. Oral Biol.* **2019**, *104*, 33–39. [CrossRef] [PubMed]
- 93. Rodríguez-Muñoz, M.; Onetti, Y.; Cortés-Montero, E.; Garzón, J.; Sánchez-Blázquez, P. Cannabidiol enhances morphine antinociception, diminishes NMDA-mediated seizures and reduces stroke damage via the sigma 1 receptor 11 Medical and Health Sciences 1109 Neurosciences 11 Medical and Health Sciences 1115 Pharmacology and Pharmaceutical Scien. *Mol. Brain* 2018, 11, 1–12.
- 94. Neelakantan, H.; Tallarida, R.J.; Reichenbach, Z.W.; Tuma, R.F.; Ward, S.J.; Walker, E.A. Distinct interactions of cannabidiol and morphine in three nociceptive behavioral models in mice. *Behav. Pharmacol.* **2015**, 26, 304–314. [CrossRef]
- 95. Crivelaro do Nascimento, G.; Ferrari, D.P.; Guimaraes, F.S.; Del Bel, E.A.; Bortolanza, M.; Ferreira, N.C., Jr. Cannabidiol increases the nociceptive threshold in a preclinical model of Parkinson's disease. *Neuropharmacology* **2020**, *163*, 107808. [CrossRef] [PubMed]
- 96. Finn, D.P.; Beckett, S.R.G.; Roe, C.H.; Madjd, A.; Fone, K.C.F.; Kendall, D.A.; Marsden, C.A.; Chapman, V. Effects of coadministration of cannabinoids and morphine on nociceptive behaviour, brain monoamines and HPA axis activity in a rat model of persistent pain. *Eur. J. Neurosci.* 2004, 19, 678–686. [CrossRef]
- 97. Cunha, J.M.; Carlini, E.A.; Pereira, A.E.; Ramos, O.L.; Pimentel, C.; Gagliardi, R.; Sanvito, W.L.; Lander, N.; Mechoulam, R. Chronic administration of cannabidiol to healthy volunteers and epileptic patients. *Pharmacology* **1980**, *21*, 175–185. [CrossRef] [PubMed]
- 98. Consroe, P.; Laguna, J.; Allender, J.; Snider, S.; Stern, L.; Sandyk, R.; Kennedy, K.; Schram, K. Controlled clinical trial of cannabidiol in Huntington's disease. *Pharmacol. Biochem. Behav.* **1991**, 40, 701–708. [CrossRef]
- 99. Wade, D.T.; Makela, P.; Robson, P.; House, H.; Bateman, C. Do cannabis-based medicinal extracts have general or specific effects on symptoms in multiple sclerosis? A double-blind, randomized, placebo-controlled study on 160 patients. *Mult. Scler.* **2004**, *10*, 434–441. [CrossRef]
- 100. Cuñetti, L.; Manzo, L.; Peyraube, R.; Arnaiz, J.; Curi, L.; Orihuela, S. Chronic Pain Treatment With Cannabidiol in Kidney Transplant Patients in Uruguay. *Transplant. Proc.* **2018**, *50*, 461–464. [CrossRef]
- 101. Taylor, L.; Gidal, B.; Blakey, G.; Tayo, B.; Morrison, G. A Phase I, Randomized, Double-Blind, Placebo-Controlled, Single Ascending Dose, Multiple Dose, and Food Effect Trial of the Safety, Tolerability and Pharmacokinetics of Highly Purified Cannabidiol in Healthy Subjects. *CNS Drugs* **2018**, *32*, 1053–1067. [CrossRef]
- 102. Devinsky, O.; Patel, A.D.; Thiele, E.A.; Wong, M.H.; Appleton, R.; Harden, C.L.; Greenwood, S.; Morrison, G.; Sommerville, K. Randomized, dose-ranging safety trial of cannabidiol in Dravet syndrome. *Neurology* **2018**, 90, e1204–e1211. [CrossRef] [PubMed]
- 103. Manini, A.F.; Yiannoulos, G.; Bergamaschi, M.M.; Hernandez, S.; Olmedo, R.; Barnes, A.J.; Winkel, G.; Sinha, R.; Jutras-Aswad, D.; Huestis, M.A.; et al. Safety and pharmacokinetics of oral Cannabidiol when administered concomitantly with intravenous Fentanyl in humans. *J. Addict. Med.* 2015, 9, 204. [CrossRef] [PubMed]
- 104. Xu, D.H.; Cullen, B.D.; Tang, M.; Fang, Y. The Effectiveness of Topical Cannabidiol Oil in Symptomatic Relief of Peripheral Neuropathy of the Lower Extremities. *Curr. Pharm. Biotechnol.* **2019**, *21*, 390–402. [CrossRef] [PubMed]
- 105. Szaflarski, J.P.; Bebin, E.M.; Cutter, G.; DeWolfe, J.; Dure, L.S.; Gaston, T.E.; Kankirawatana, P.; Liu, Y.; Singh, R.; Standaert, D.G.; et al. Cannabidiol improves frequency and severity of seizures and reduces adverse events in an open-label add-on prospective study. *Epilepsy Behav.* 2018, 87, 131–136. [CrossRef]

- 106. Devinsky, O.; Patel, A.D.; Cross, J.H.; Villanueva, V.; Wirrell, E.C.; Privitera, M.; Greenwood, S.M.; Roberts, C.; Checketts, D.; VanLandingham, K.E.; et al. Effect of cannabidiol on drop seizures in the lennox–gastaut syndrome. *N. Engl. J. Med.* **2018**, *378*, 1888–1897. [CrossRef]
- 107. Thiele, E.A.; Marsh, E.D.; French, J.A.; Mazurkiewicz, M.B.; Benbadis, S.R.; Joshi, C.; Lyons, P.D.; Taylor, A.; Roberts, C.; Sommerville, K.; et al. Cannabidiol in patients with seizures associated with Lennox-Gastaut syndrome (GWPCARE4): A randomised, double-blind, placebo-controlled phase 3 trial. *Lancet* 2018, 391, 1085–1096. [CrossRef]
- 108. Laux, L.C.; Bebin, E.M.; Checketts, D.; Chez, M.; Flamini, R.; Marsh, E.D.; Miller, I.; Nichol, K.; Park, Y.; Segal, E.; et al. Long-term safety and efficacy of cannabidiol in children and adults with treatmentresistant Lennox-Gastaut syndrome or Dravet syndrome: Expanded access program results. *Epilepsy Res.* 2019, 154, 13–20. [CrossRef]
- 109. Devinsky, O.; Cross, J.H.; Laux, L.; Marsh, E.; Miller, I.; Nabbout, R.; Scheffer, I.E.; Thiele, E.A.; Wright, S. Trial of cannabidiol for drug-resistant seizures in the dravet syndrome. *N. Engl. J. Med.* **2017**, *376*, 2011–2020. [CrossRef]
- 110. Capano, A.; Weaver, R.; Burkman, E. Evaluation of the effects of CBD hemp extract on opioid use and quality of life indicators in chronic pain patients: A prospective cohort study. *Postgrad. Med.* **2020**, *132*, 56–61. [CrossRef]
- 111. Van De Donk, T.; Niesters, M.; Kowal, M.A.; Olofsen, E.; Dahan, A.; Van Velzen, M. An experimental randomized study on the analgesic effects of pharmaceutical-grade cannabis in chronic pain patients with fibromyalgia. *Pain* **2019**, *160*, 860–869. [CrossRef]
- 112. Hoggart, B.; Ratcliffe, S.; Ehler, E.; Simpson, K.H.; Hovorka, J.; Lejčko, J.; Taylor, L.; Lauder, H.; Serpell, M. A multicentre, open-label, follow-on study to assess the long-term maintenance of effect, tolerance and safety of THC/CBD oromucosal spray in the management of neuropathic pain. *J. Neurol.* **2015**, 262, 27–40. [CrossRef] [PubMed]
- 113. Serpell, M.; Ratcliffe, S.; Hovorka, J.; Schofield, M.; Taylor, L.; Lauder, H.; Ehler, E. A double-blind, randomized, placebo-controlled, parallel group study of THC/CBD spray in peripheral neuropathic pain treatment. *Eur. J. Pain* **2014**, *18*, 999–1012. [CrossRef] [PubMed]
- 114. Johnson, J.R.; Burnell-Nugent, M.; Lossignol, D.; Ganae-Motan, E.D.; Potts, R.; Fallon, M.T. Multicenter, Double-Blind, Randomized, Placebo-Controlled, Parallel-Group Study of the Efficacy, Safety, and Tolerability of THC:CBD Extract and THC Extract in Patients with Intractable Cancer-Related Pain. *J. Pain Symptom Manag.* 2010, 39, 167–179. [CrossRef] [PubMed]
- 115. Johnson, J.R.; Lossignol, D.; Burnell-Nugent, M.; Fallon, M.T. An open-label extension study to investigate the long-term safety and tolerability of THC/CBD oromucosal spray and oromucosal THC spray in patients with terminal cancer-related pain refractory to strong opioid analgesics. *J. Pain Symptom Manag.* 2013, 46, 207–218. [CrossRef]
- 116. Russo, M.; Naro, A.; Leo, A.; Sessa, E.; D'Aleo, G.; Bramanti, P.; Calabrò, R.S. Evaluating sativex<sup>®</sup> in neuropathic pain management: A clinical and neurophysiological assessment in multiple sclerosis. *Pain Med.* **2016**, *17*, 1145–1154. [CrossRef]
- 117. Portenoy, R.K.; Ganae-Motan, E.D.; Allende, S.; Yanagihara, R.; Shaiova, L.; Weinstein, S.; McQuade, R.; Wright, S.; Fallon, M.T. Nabiximols for opioid-treated cancer patients with poorly-controlled chronic pain: A randomized, placebo-controlled, graded-dose trial. *J. Pain* 2012, *13*, 438–449. [CrossRef]
- 118. Nurmikko, T.J.; Serpell, M.G.; Hoggart, B.; Toomey, P.J.; Morlion, B.J.; Haines, D. Sativex successfully treats neuropathic pain characterised by allodynia: A randomised, double-blind, placebo-controlled clinical trial. *Pain* 2007, 133, 210–220. [CrossRef]
- 119. Blake, D.R.; Robson, P.; Ho, M.; Jubb, R.W.; McCabe, C.S. Preliminary assessment of the efficacy, tolerability and safety of a cannabis-based medicine (Sativex) in the treatment of pain caused by rheumatoid arthritis. *Rheumatology* **2006**, *45*, 50–52. [CrossRef]
- 120. Lichtman, A.H.; Lux, E.A.; McQuade, R.; Rossetti, S.; Sanchez, R.; Sun, W.; Wright, S.; Kornyeyeva, E.; Fallon, M.T. Results of a Double-Blind, Randomized, Placebo-Controlled Study of Nabiximols Oromucosal Spray as an Adjunctive Therapy in Advanced Cancer Patients with Chronic Uncontrolled Pain. *J. Pain Symptom Manag.* 2018, 55, 179–188. [CrossRef]

- 121. Lynch, M.E.; Cesar-Rittenberg, P.; Hohmann, A.G. A double-blind, placebo-controlled, crossover pilot trial with extension using an oral mucosal cannabinoid extract for treatment of chemotherapy-induced neuropathic pain. *J. Pain Symptom Manag.* **2014**, *47*, 166–173. [CrossRef]
- 122. Santoro, M.; Mirabella, M.; De Fino, C.; Bianco, A.; Lucchini, M.; Losavio, F.; Sabino, A.; Nociti, V. Sativex<sup>®</sup> effects on promoter methylation and on CNR1/CNR2 expression in peripheral blood mononuclear cells of progressive multiple sclerosis patients. *J. Neurol. Sci.* **2017**, *379*, 298–303. [CrossRef] [PubMed]
- 123. Martin-Santos, R.A.; Crippa, J.; Batalla, A.; Bhattacharyya, S.; Atakan, Z.; Borgwardt, S.; Allen, P.; Seal, M.; Langohr, K.; Farre, M.; et al. Acute Effects of a Single, Oral dose of d9-tetrahydrocannabinol (THC) and Cannabidiol (CBD) Administration in Healthy Volunteers. *Curr. Pharm. Des.* **2012**, *18*, 4966–4979. [CrossRef] [PubMed]
- 124. Schoedel, K.A.; Szeto, I.; Setnik, B.; Sellers, E.M.; Levy-Cooperman, N.; Mills, C.; Etges, T.; Sommerville, K. Abuse potential assessment of cannabidiol (CBD) in recreational polydrug users: A randomized, double-blind, controlled trial. *Epilepsy Behav.* 2018, 88, 162–171. [CrossRef] [PubMed]
- 125. Ignatowska-Jankowska, B.; Jankowski, M.; Glac, W.; Swiergiel, A.H. Cannabidiol-induced lymphopenia does not involve NKT and NK cells. *J. Physiol. Pharmacol.* **2009**, *60*, 99–103. [PubMed]
- 126. Calvey, T.N.; Williams, N.E. *Principles and Practice of Pharmacology for Anaesthetists*; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2008.

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